

DEVELOPMENT OF DIAGNOSTICS FOR SWEETPOTATO FEATHERY MOTTLE VIRUS

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INTRODUCTION

Sweet potato (*Ipomoea batatas* L.) is an important starchy tuberous root crop after potato and cassava, that feeds millions of people in developing countries.

Productivity of sweet potato is very low in India (8 tonnes/ ha)

Constraints

Sweet potato weevil and viral diseases are the major biotic constraints

Viral Diseases : SPFMV is widely prevalent

Production of healthy quality planting material is one of major mandate of CTCRI

OBJECTIVES

- Development of PCR based protocols for detection of SPFMV
- Cloning and sequencing of coat protein gene of SPFMV occurring in India

Symptoms of feathery mottle disease



Contd..

Symptoms of feathery mottle disease



Contd..



Symptoms of feathery mottle disease

Transmission to tobacco



Nicotiana benthamiana

Nicotiana tabacum

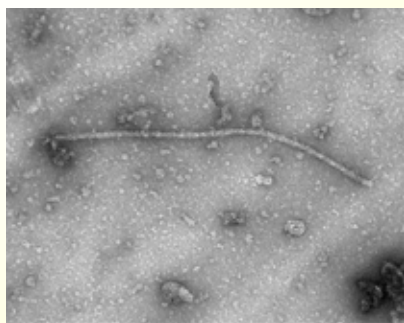
Host plants	Symptoms	Symptoms
<i>Nicotina tabacum</i>	8/14	Mosaic vein clearing and
<i>Nicotiana benthamiana</i>	4/12	Chlorotic spot and leaf distruption
<i>Ipomoea nil</i>	11/15	Chlorotic spot and vein clearing
<i>Ipomoea setosa</i>	10/15	Vein clearing ,leaf distortion, mosaic

Transmission to *Ipomoea setosa*



Graft transmission

Sap inoculation

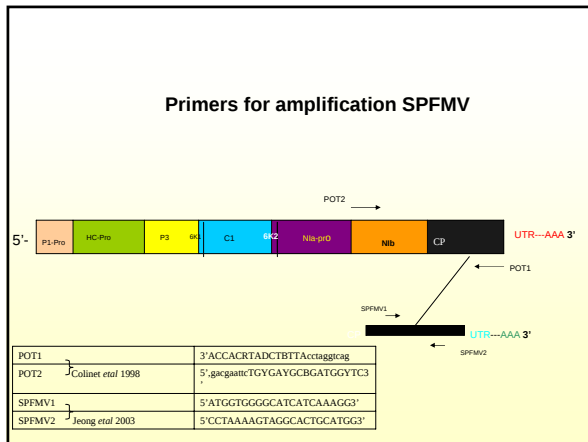


Flexuous SPFMV particles

Structural organisation of SPFMV

PROTEIN	POSSIBLE FUNCTION(S)
P1	Proteinase; Cell-to-cell movement (speculation); Aphid mediated transmission;
HC-Pro	Proteinase; Cell-to-cell movement (speculation); Unknown (possible role in replication)
P3	Unknown (possible role in replication)
CI	Genome replication (RNA helicase); Membrane attachment; Nucleic acid stimulated ATPase activity; Cell-to-cell movement (speculation).
CP	RNA encapsidation; Involved in vector transmission; Cell-to-cell movement.
N1a-VPg	Genome replication (Primer for initiation of RNA synthesis).
N1a-Pro	Major Proteinase
N1b	Genome replication (RNA-dependent RNA polymerase (RdRp)).
SK1 & SK2	Unknown, but possible roles in: - RNA replication; - Regulatory function inhibiting N1a nuclear translocation; - Membrane anchoring of replication machinery

5' VPg 200 400 600 800 1000 1200 1400 1600 1800 2000 2200 2400 2600 2800 3000 3200 3400 3600 3800 4000 4200 4400 4600 4800 5000 -AAA3'



RT PCR

RNA	100-150ng
10x buffer	2µl
dNTP Mix	~.25µl
Cmaster RT enzyme	~.25µl
CmasterTaq enzyme	~.25µl
Rnase inhibitor	0.2µl
Nuclease free water	-
Total	20µl

SPFMV1 & SPFMV2

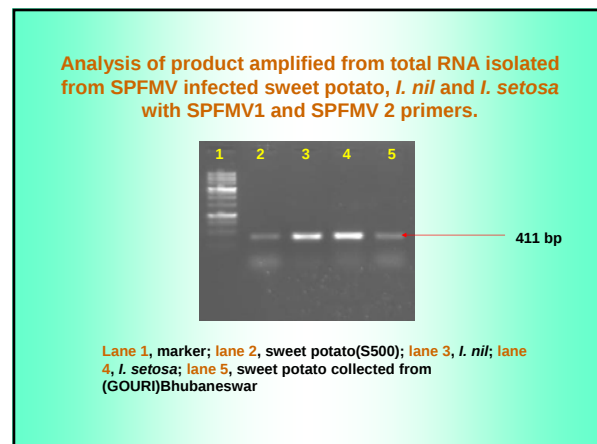
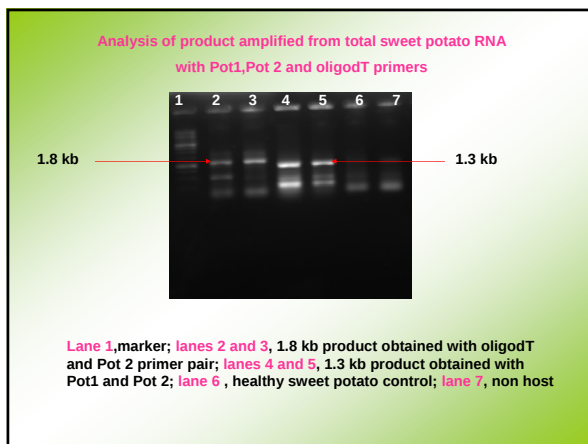
42°C -45 minutes
96 °C -2 minutes
96 °C -30seconds
63 °C -30 seconds
72 °C -1 minute
72 °C -7 minutes

} 40 cycles

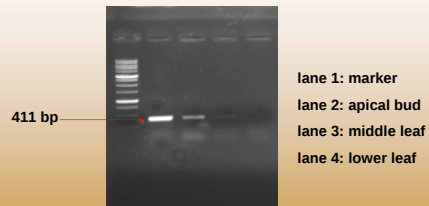
POT 1& POT 2

42°C -45 minutes
92 °C -2 minutes
94 °C -30seconds
42°C -1min
72°C - 90seconds
94-30seconds
60-60 seconds
72°C -90seconds
72 °C -5 minute

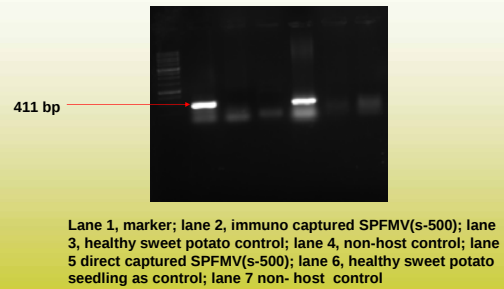
} 35 cycles



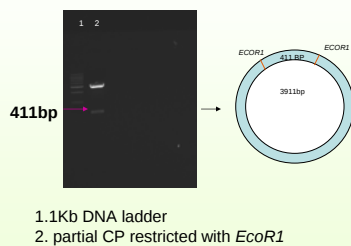
Detection of SPFMV from different parts of infected sweet potato plants with SPFMV 1 and SPFMV 2 primers



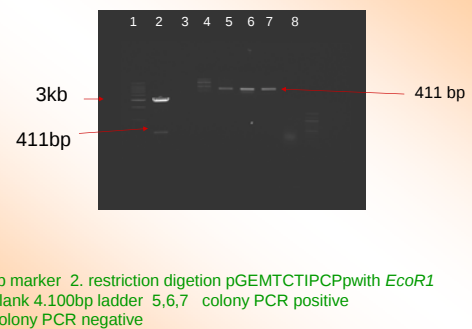
Detection of SPFMV in sweet potato by IC-RT-PCR and direct tube capture RT-PCR.

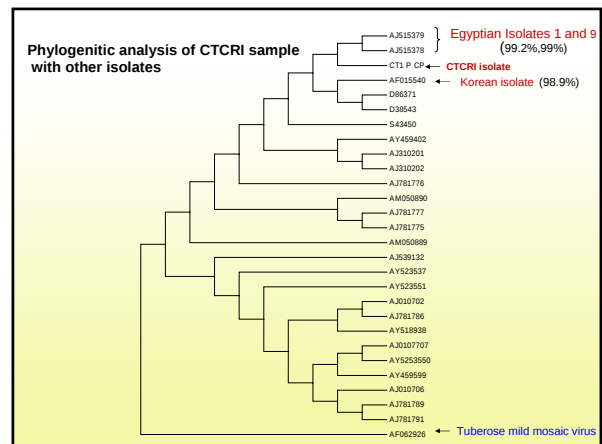
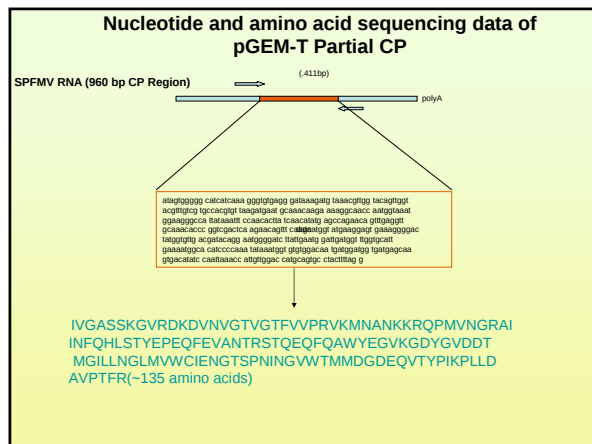


DIAGRAMATIC REPRESENTATION
pGEM-T Partial CP



CLONE CONFIRMATION WITH RESTRICTION
DIGESTION AND COLONY PCR





Plants tested by PCR	POT 1&POT 2 PRIMER	SPFMV 1 & SPFMV 2 PRIMER
S-500	+	+
S-316	-	+
S-211	-	+
S-629	+	+
S1070	+	+
S-315	+	+
S-306	-	+
S-406	-	-
S-382	+	+
S-465	-	+
S-658	+	+
S-492	+	+
S-1114	-	+
S-703	-	+
S-645	+	+
S-101	-	-
S-343	-	+
Lnil	+	+
Isetosa	+	+

Detection of SPFMV in sweet potato clones by RT-PCR using Pot 1 and Pot 2 primers and SPFMV 1 and SPFMV 2 primers

Conclusion

- SPFMV could be detected using potyvirus specific degenerate(POT1&POT2) primers and gene specific SPFMV1& SPFMV2 primers
- IC RT- PCR and tube capture PCR was standardized using gene specific primers
- Partial SPFMV CP was amplified and cloned in pGEM-T vector
- Phylogenetic analysis showed that pGEM-T partial CP having 99% similarity to Egyptian isolates.

